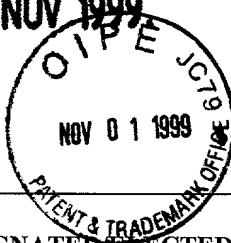


U.S. Application No.

International Application No.  
PCT/EP98/02590

Date: November 1, 1999

Attorney Docket No.  
VANM131.001APC  
Page 1TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US)  
CONCERNING A FILING UNDER 35 USC 371

International Application No.: PCT/EP98/02590  
 International Filing Date: May 4, 1998  
 Priority Date Claimed: April 30, 1997  
 Title of Invention: INHIBITORS OF CELLULOLYTIC, XYLANOLYTIC AND B-GLUCANOLYTIC ENZYMES  
 Applicant(s) for DO/EO/US: Debysy et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1.  This is a **FIRST** submission of items concerning a filing under 35 USC 371.
2.  This express request to begin national examination procedures (35 USC 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 USC 371(b) and PCT Articles 22 and 39(1).
3.  A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
4.  A copy of the International Application as filed (35 USC 371(c)(2))
  - a)  is transmitted herewith (required only if not transmitted by the International Bureau).
  - b)  has been transmitted by the International Bureau.
  - c)  is not required, as the application was filed in the United States Receiving Office (RO/US).
5.  A translation of the International Application into English (35 USC 371(c)(2)).
6.  Amendments to the claims of the International Application under PCT Article 19 (35 USC 371(c)(3))
  - a)  are transmitted herewith (required only if not transmitted by the International Bureau).
  - b)  have been transmitted by the International Bureau.
  - c)  have not been made; however, the time limit for making such amendments has NOT expired.
  - d)  have not been made and will not be made.
7.  A copy of the International Preliminary Examination Report with any annexes thereto, such as any amendments made under PCT Article 34.
8.  A **FIRST** preliminary amendment.
9.  International Application as published.
10.  Diskette containing machine readable form of Sequence Listing.
11.  International Search Report.
12.  A return prepaid postcard.

U.S. Application No.  
**09/403625**  
 Date: November 1, 1999

International Application No.  
 PCT/EP98/02590

Attorney Docket No.  
 VANM131.001APC  
 Page 2

The following fees are submitted:

FEES				
BASIC FEE				\$840
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	
Total Claims	47 - 20 =	27 x	\$18	\$486
Independent Claims	1 - 3 =	0 x	\$78	\$0
Multiple dependent claims(s) (if applicable)			\$260	\$0
<b>TOTAL OF ABOVE CALCULATIONS</b>				<b>\$1326</b>

13.  A check in the amount of \$840 to cover the basic fee only is enclosed.

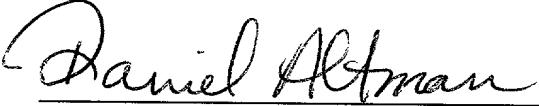
14.  The fees for excess claims and for later submission of the signed oath or declaration set forth in 37 CFR 1.492(e) will be paid upon submission of the declaration.

15.  The Commissioner is hereby authorized to charge only those additional fees which may be required to avoid abandonment of the application, or credit any overpayment to Deposit Account No. 11-1410. A duplicate copy of this sheet is enclosed.

**NOTE:** Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

KNOBBE, MARTENS, OLSON & BEAR, LLP  
 Customer No. 20,995

  
 Daniel E. Altman  
 Registration No. 34,115

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09/403625

430 Rec'd PCT/PIO 01 NOV 1999

VANM131.001APC

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant	:	Debyser, et al.	)	Group Art Unit Unknown
			)	)
Intl. Appl. No.	:	PCT/EP98/02590	)	)
			)	)
Intl. Filed	:	May 4, 1998	)	)
			)	)
For	:	INHIBITORS OF CELLULOLYTIC, XYLANOLYTIC AND $\beta$ - GLUCANOLYTIC ENZYMES	)	)
			)	)
Examiner	:	Unknown	)	)

PRELIMINARY AMENDMENT  
INCLUDING  
SEQUENCE LISTING

Assistant Commissioner for Patents  
Washington, D.C. 20231

Dear Sir:

Preliminary to examination on the merits, please amend the above-referenced application as follows:

IN THE SPECIFICATION:

On page 1 of the Specification, after the Title of the Invention ending on line 8 and before "Field of the Invention" on line 10, please insert: --This is the U.S. national phase under 35 U.S.C. § 371 of International Application PCT/EP98/02590, filed May 4, 1998.--

On page 27, please delete "Claims" and substitute --WHAT IS CLAIMED IS:--

IN THE CLAIMS:

**Please amend the claims as follows:**

1. **(Amended)** A proteinic or glycoproteinic inhibitor of an enzyme selected from the group consisting of cellulolytic, xylanolytic and [or]  $\beta$ -glucanolytic enzymes.

2. (Amended) The [I]inhibitor [as in]of claim 1, [characterized in that]wherein said inhibitor inhibits enzymes selected from the group consisting of: cellulase, endoxylanase,  $\beta$ -glucanase,  $\beta$ -xylosidase, and  $\alpha$ -L-arabinofuranosidase [and/or other cellulose, xylan, arabinoxylan or  $\beta$ -glucan degrading enzymes].

3. (Amended) The [I]inhibitor [as in]of claim 1 [or 2, characterized in that it] wherein said inhibitor is obtainable from plant material or fractions thereof.

4. (Amended) The [I]nhibitor [as in]of claim 3, [characterized in that] wherein said plant material is selected from the group consisting of cereals, cereal grains from wheat, [or] cereal flours from wheat, durum wheat, rye, triticale, barley, sorghum, oats, maize [or]and rice.

5. (Amended) The [I]inhibitor [as in]of claim 1 [or 2, characterized in that it] wherein said inhibitor is obtainable from micro-organisms or fractions thereof.

6 (Amended) The [I]inhibitor [as in any of the] of claim[s] 1 [-5, characterized in that it] wherein said inhibitor is a xylanase inhibitor.

7. (Amended) The [I]inhibitor [as in]of claim 6, [characterized in that it] wherein said inhibitor is [a] water-soluble [species].

8. (Amended) The [I]inhibitor [as in]of claim 7 comprising[having a marker whose]an amino acid sequence [has more than]at least 70% [homology with] homologous to [SEQ ID No. 1 and/or SEQ ID No. 2]SEQ ID NO:1 or SEQ ID NO:2.

9. (Amended) The [I]inhibitor [as in]of claim 8, [characterized in that] wherein the [marker]amino acid sequence is the N-terminal amino acid sequence of the protein or glycoprotein.

10. (Amended) The [I]inhibitor [as in]of claim 8 [or 9 having] comprising [a marker for which]an amino acid sequence [has more than]at least 85% [homology with]homologous to [SEQ ID No. 1 and/or SEQ ID No. 2]SEQ ID NO:1 or SEQ ID NO:2.

11. (Amended) The [I]inhibitor [as in]of claim 10, [characterized in that] wherein the [marker]amino acid sequence is the N-terminal amino acid sequence of the protein or glycoprotein.

12. (Amended) The [I]inhibitor [as in]of claim 7, comprising[having a marker whose amino acid sequence is identical to SEQ ID No. 1 and/or SEQ ID No. 2]SEQ ID NO:1 or SEQ ID NO:2.

13. (Amended) The [I]inhibitor [as in]of claim 12, [characterized in that] wherein [the marker] SEQ ID NO:1 or SEQ ID NO:2 is the N-terminal amino acid sequence of the protein or glycoprotein.

14. (Amended) The [I]inhibitor [as in any of the] of claim[s] 7 [to 13, characterized in that] wherein said protein or glycoprotein is selected from the group [comprising] consisting of proteins or glycoproteins having a molecular weight [typically] between approximately 40 kDa and 43 kDa, proteins or glycoproteins having a molecular weight [typically] of approximately 30 kDa and proteins or glycoproteins having a molecular weight of [typically] approximately 10 kDa.

15. (Amended) The [I]inhibitor [as in any of the] of claim[s] 7 [to 14, characterized in that] wherein said protein or glycoprotein [typically] has a molecular weight between approximately 40 kDa and 43 kDa and has a pI of greater than about 7.

16. (Amended) A [M]method for obtaining the inhibitor [as in]of [any of the] claim[s] 1 [to 15] from [possibly genetically modified] micro-organisms, plants or plant materials, [wherein said micro-organisms, plants or plant materials are] comprising subject[ed]ing said micro-organisms, plants, or plant materials to one or more extraction and/or fractionation steps.

17. (Amended) A [M]method for obtaining the inhibitor [according to any of the] of claim[s] 1 [to 15, wherein] comprising genetically modifying micro-organisms, plants or plant materials [are genetically modified] by the introduction of a genetic material encoding said inhibitor into the micro-organisms, plants or plant materials.

18. (Amended) A [P]process for transforming micro-organisms, plants or plant materials, [wherein] comprising reducing the activity of the inhibitor [according to any of the] of claim[s] 1 [to 15 is reduced].

19. (Amended) The [P]process according to claim 18, [characterized in that] wherein the [reduced] activity of the inhibitor [according to the invention is obtained] is reduced by reduction of its expression.

20. (Amended) The [P]process according to claim 18 [or 19, characterized in that] wherein the activity of the inhibitor is reduced by blocking the inhibitor function.

21. (Amended) A [P]rocess for transforming micro-organisms, plants or plant materials, [wherein]comprising increasing the activity of the inhibitor according to [any of the] claim[s] 1 [to 15 is increased].

22. (Amended) The [P]rocess according to claim 21, [characterized in that] wherein the [increased] activity of the inhibitor [according to the invention is obtained] is increased by an increase of its expression.

23. (Amended) The [P]rocess according to claim 21 [or 22, characterized in that] wherein the activity of the inhibitor is increased by activating the inhibitor function.

24. (Amended) A [M]icro-organism[s], plants or plant material[s] obtained by the method according to [any of the preceding]claim[s] 17 [to 23].

25. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants and/or the plant materials according to claim 24]A method for improving the production of beer or the malting of cereals [such as barley, sorghum and wheat and/or the production of beer] comprising; adding the inhibitor of Claim 1 to the cereal malt.

26. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 24, the micro-organisms, the plants or the plant materials according to claim 24]A method for improving the production and/or quality of baked or extruded cereal products [chosen among the group consisting of straight dough, sponge dough, Chorleywood bread, breakfast cereals, biscuits, pasta and noodles]comprising adding the inhibitor of claim 1, before baking, to the baked or extruded cereal product.

27. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants or the plant materials according to claim 24]A method for improving animal feedstuff efficiency comprising adding the inhibitor of claim 1 to the animal feedstuff.

28. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 15, the micro-organisms, the plants or the plant materials according to claim 24]A method for improving the production of starch derived products selected from the group consisting of: syrups, sorbitol, xylose and[or] xylitol; comprising adding the inhibitor of claim 1 to the starch derived products.

29. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants or the plant materials according to claim 24] A method for wheat gluten-starch separation and production comprising adding the inhibitor of claim 1 to the wheat gluten-starch mixture.

30. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants or the plant materials according to claim 24] A method for improving maize processing comprising adding the inhibitor of claim 1 to the maize.

31. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants or the plant materials according to claim 24] A method for improving plant disease resistance comprising adding the inhibitor of claim 1 to the plant.

32. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants or the plant materials according to claim 24] A method for improving nutraceutical and/or pharmaceutical applications comprising adding the inhibitor of claim 1 to the nutraceutical and/or pharmaceutical.

33. (Amended) [Use of the inhibitor according to any of the preceding claims 1 to 15 or obtained by the method of claim 16, the micro-organisms, the plants or the plant materials according to claim 24] A method for improving paper and pulp technologies comprising adding the inhibitor of Claim 1 to said paper and pulp mixtures during production of said paper and pulp.

**Please add the following claims:**

34. The method of Claim 16 wherein said micro-organisms, plants or plant materials are genetically modified.

35. A method for improving the production of beer or the malting of cereals comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24 to said cereal malt.

36. The method of Claim 35 wherein said cereals are selected from the group consisting of: barley, sorghum and wheat.

37. The method of Claim 25 wherein said cereals are selected from the group consisting of: barley, sorghum and wheat.

38. A method for improving the production and/or quality of baked or extruded cereal products comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, before baking, to the baked or extruded cereal product.

39. The method of Claim 26 wherein said cereal products are selected from the group consisting of straight dough, sponge dough, Chorleywood bread, breakfast cereals, biscuits, pasta and noodles.

40. A method for improving animal feedstuff efficiency comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24 to the animal feedstuff.

41. A method for improving the production of starch derived products selected from the group consisting of: syrups, sorbitol, xylose and xylitol: comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, to the starch derived products.

42. A method for wheat gluten-starch separation and production comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, to the wheat gluten-starch mixture.

43. A method for improving maize processing comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, to the maize.

44. A method for improving plant disease resistance comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, to the plant.

45. A method for improving nutraceutical and/or pharmaceutical applications comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, to the nutraceutical and/or pharmaceutical.

46. A method for improving paper and pulp technologies comprising adding the inhibitor obtained from the microorganisms, the plants or the plant material of claim 24, to said paper and pulp mixtures during production of said paper and pulp.

47. The method of claim 31 wherein said inhibitor is added by transforming the plant cells with a vector expressing said inhibitor.

**IN THE SEQUENCE LISTING:**

Please insert the attached Sequence Listing page 1.

**REMARKS**

Intl. Appl. No. : PCT/EP98/02590  
Intl. Filed : May 4, 1998

The amendments made herein conform the application to practice in the United States Patent and Trademark Office. Claims 1-33 have been amended. Claims 33-47 have been added. The amended and added claims are substantially based on the claims as originally filed. Thus, none of the claims added by this Preliminary Amendment introduce new matter into the application. The specification has been amended to include the International priority application. A Sequence Listing in both paper and computer-readable formats is provided herewith.

**VERIFICATION UNDER 37 C.F.R. §1.821(f) & (g)**

The polypeptide sequence appearing in the attached Sequence Listing was included in the application as filed. Pursuant to 37 C.F.R. §1.821(g), no new matter is being added herewith. As required under 37 C.F.R. §1.821(f), I hereby verify that the data on the enclosed disk and the paper copies of the Sequence Listing are identical.

**CONCLUSION**

Entry of this Preliminary Amendment and examination on the merits are respectfully requested.

Respectfully submitted,

KNOBBE, MARTENS, OLSON & BEAR, LLP

Dated: 1 Nov. 1999

By:



Daniel E. Altman  
Registration No. 34,115  
Attorney of Record  
620 Newport Center Drive  
Sixteenth Floor  
Newport Beach, CA 92660  
(949) 760-0404

SEQUENCE LISTING

<110> Debyser, Winok  
Delcour, Jan

<120> INHIBITORS OF CELLULOLYTIC, XYLANOLYTIC  
AND BETA-GLUCANOLYTIC ENZYMES.

<130> VANM131.001APC

<150> PCT/EP98/02590  
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protein or glycoprotein.

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<223> Xaa = Any Amino Acid, preferably Asp.

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<221> VARIANT  
<222> (1)...(17)  
<223> Xaa = Any Amino Acid, but the first Xaa is  
preferably Ser, Phe, or Gly.

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1 5 10 15  
Val

INHIBITORS OF CELLULLOLYTIC, XYLANOLYTIC AND  $\beta$ -GLUCANOLYTIC ENZYMES

10 Field of the invention

This invention relates to an inhibitor of cellulolytic, xylanolytic and/or  $\beta$ -glucanolytic enzymes (sometimes also referred to as cellulases, pentosanases and/or hemicellulases) especially an inhibitor of pentosan degrading enzymes such as endoxylanase (such as EC: 3.2.1.8),  $\beta$ -xylosidase (such as EC: 3.2.1.37), and  $\alpha$ -L-arabinofuranosidase (such as EC: 3.2.1.55), to inhibitors of cellulase (such as EC: 3.2.1.4),  $\beta$ -glucanase (such as EC: 3.2.3.73 or EC: 3.2.1.6), and to inhibitors of other 20 xylan, arabinoxylan and  $\beta$ -glucan degrading enzymes, which are present in micro-organisms, plants, plant materials or fractions thereof, (such as cereals, cereal grains, cereal flours or fractions thereof).

The present invention is also related to a 25 method for obtaining said inhibitor, as well as to the use of said inhibitor in different areas of food, feed and/or beverage technologies, such as malting and brewing, the production of animal feedstuffs such as to increase their conversion, the production of baked and/or extruded cereal 30 products such as straight dough, sponge and dough and Chorleywood breads, breakfast cereals, different types of

biscuits, pasta and noodles, the production of starch derived syrups, sorbitol, xylose and/or xylitol, the wheat gluten-starch separation industry, maize processing, the improvement of plant disease resistance, in nutraceutical 5 or pharmaceutical applications such as maintaining the structure of dietary fiber material, and in the field of paper and pulp technologies.

Background of the invention

10 Apart from barley malt, unmalted cereals such as wheat are commonly used in beer production (Pierce, J.S., *Proceedings of the European Brewery Convention Congress, Madrid, 1987*, 445). Unmalted wheat (40-50%) is e.g. used for the production of Belgian white (wheat) 15 beers.

Although barley and wheat endosperm cell walls contain 20 and 70% (w/w) arabinoxylan respectively (Ballance, G.M., & Manners, D.J., *Carbohydrate Research*, 1978, 61,107; Fincher, G.B., & Stone, B.A. In: *Advances in 20 Cereal Science and Technology*, Vol. VIII. Y. Pomeranz, (Ed), Am. Assoc. Cereal Chem., St. Paul (MN), 1986, 207), their total arabinoxylan content is comparable, i.e. 2.8 to 7.1% (w/w) for barley and 3.6 to 7.1% (w/w) for wheat (Henry, J., *Journal of the Science of Food and Agriculture*, 25 1985, 36, 1243; Hashimoto, S., Shogren, M.D. & Pomeranz, Y., *Cereal Chemistry*, 1987, 64, 30).

The grains also contain comparable levels of water-extractable arabinoxylan, i.e. 0.24 to 0.80% (w/w) for barley and 0.25 to 1.18% for wheat (Henry, J., *Journal 30 of the Science of Food and Agriculture*, 1985, 36, 1243; Hashimoto, S., Shogren, M.D. & Pomeranz, Y., *Cereal Chemistry*, 1987, 64, 30; Åman, P., & Hesselman, K., *Swedish*

Journal of Agricultural Research, 1984, 14, 135; Girhammer, U., & Nair, B.M., Food Hydrocolloids, 1992, 6, 285). Furthermore, barley and wheat endosperm cell walls contain 70 and 20%  $\beta$ -glucan respectively (Ballance, G.M., & 5 Manners, D.J., Carbohydrate Research, 1978, 61, 107; Fincher, G.B., & Stone, B.A. In: Advances in Cereal Sciences and Technology, Vol. VIII. Y. Pomeranz, (Ed), Am. Assoc. Cereal Chem., St. Paul (MN), 1986, 207).

Barley grains contain 1.7 to 4.1% (w/w)

10 water-extractable and 3.6 to 6.4% (w/w) total  $\beta$ -glucan (Anderson, M.A., Cook, J.A., & Stone, B.A., Journal of the Institute of Brewing, 1978, 84, 233-239; Henry, J., Journal of the Science of Food and Agriculture, 1985, 36, 1243). Wheat grains contain 0.1 to 0.8% (w/w) water-extractable 15 and 0.6 to 1.4% (w/w) total  $\beta$ -glucan (Anderson, M.A., Cook, J.A., & Stone, B.A., Journal of the Institute of Brewing, 1978, 84, 233-239; Henry, J., Journal of the Science of Food and Agriculture, 1985, 36, 1243). As in wheat only low levels of arabinoxylan (Cleemput, G., Bleukx, W., van Oort, 20 M., Hessing, M. & Delcour, J.A., Journal of Cereal Science, 1995, 22, 139) and  $\beta$ -glucan degrading enzyme activities can be measured, the barley malt must be mostly responsible for wheat and malt arabinoxylan and  $\beta$ -glucan hydrolysis during brewing.

25 Efficient hydrolysis of arabinoxylans and  $\beta$ -glucan is important because such compounds can be involved in production problems such as wort viscosity (Ducroo, P. & Frelon, P.G., Proceedings of the European Brewery Convention Congress, Zurich, 1989, 445; Viëtor, R.J. & 30 Voragen, A.G.J., Journal of the Institute of Brewing, 1993, 99, 243) and filterability and haze formation (Coote, N. &

Kirsop, B.H. 1976., *Journal of the Institute of Brewing*, 1976, 82, 34; Izawa, M., Kano, Y. & Kanimura, M. 1991. *Proceedings Aviemore Conference on Malting, Brewing and Distilling*, 1990, 427).

5 In other areas efficient hydrolysis of xylans and/or arabinoxylans is highly desirable as well. Examples include rye and wheat breadmaking processes, paper and pulp technologies. It follows that a lot of research efforts have been devoted to the (potential) applications of xylan  
10 and/or arabinoxylan hydrolysis enzymes due to their applications as described above.

#### Summary of the invention

The present invention concerns an inhibitor  
15 of cellulolytic, xylanolytic and/or  $\beta$ -glucanolytic enzymes, preferably an inhibitor of endoxylanase, of  $\beta$ -glucanase, of  $\beta$ -xylosidase, of  $\alpha$ -L-arabinofuranosidase, and of other xylan, arabinoxylan and  $\beta$ -glucan degrading enzymes preferably obtained from micro-organisms, plants, plant  
20 materials or fractions thereof (such as cereals, cereal grains, cereal germs or fractions thereof, cereal flours or fractions thereof).

"An inhibitor of an enzyme" means a molecule which is able to inhibit partially or totally the activity  
25 of said enzyme. In irreversible inhibition, the inhibitor is covalently linked to the enzyme or bound so tightly that its dissociation from the enzyme is very slow. In this case, the inhibitor usually mimicks the normal substrate of said enzyme in a cross-linking reaction. In contrast,  
30 reversible inhibition may be characterised by a rapid equilibrium between the enzyme and the inhibitor. A

competitive inhibitor prevents the substrate from binding to the active site and may reduce the reaction rate by diminishing the proportion of enzyme molecules that are bound to substrate. In non-competitive inhibition, the inhibitor may decrease the turnover number. Competitive inhibition can be distinguished from non-competitive inhibition by determining whether the inhibition can be overcome by raising the substrate concentration. Inhibitors isolated from a specific biological species and that are of proteinaceous or glycoproteinaceous nature can be active against enzymes of the same species (i.e. endogeneous enzymes) and/or against enzymes of different species (i.e. exogeneous enzymes).

Advantageously the inhibitor of the invention can be produced by micro-organisms or may be present in various extraction media from micro-organisms or plant material, such as cereals or fractions thereof, such as cereal grains or fractions thereof, such as cereal germs or fractions thereof, such as cereal flours or fractions thereof, such as from wheat, durum wheat, rye, triticale, barley, sorghum, oats, maize and/or rice, from which it can be obtained by the methods well known by the person skilled in the art. According to a preferred embodiment of the present invention, the inhibitor is a xylanase inhibitor which is typically water-soluble alkaline proteinaceous species, having a pI (i.e. -log of the isoelectric point) of greater than about 7.0. The xylanase inhibitor molecular weight as determined by SDS-page is typically 40-43 kDa. Following reduction with  $\beta$ -mercaptoethanol three SDS-page protein bands are found with SDS-page molecular weights of ca. 40-43 kDa, ca. 30 kDa, and ca. 10 kDa. The N-terminal sequence of the 40-43 kDa protein or glycoprotein has not

been described until now and is typically as follows: SEQ ID No. 1: Lys-Gly-Leu-Pro-Val-Leu-Ala-Pro-Val-Thr-Lys-Xaa-Thr-Ala, wherein Xaa being preferably Asp. The 30 kDa band has the above described typical N-terminal amino acid SEQ 5 ID No.1, while the N-terminal amino acid sequence of the 10 kDa band is typically as follows: SEQ ID No. 2: Xaa-Ala-Pro-Val-Ala-Lys-Met-Val-Leu-Pro-Val-Ala-Met-Lys-Glu-Xaa-Val, wherein the first Xaa being preferably Ser, Phe, or Gly, and wherein the second Xaa is unidentified. This 10 sequence has not been described before.

Therefore, the present invention is also related to an inhibitor with an SDS-page molecular weight of typically 40-43 kDa being a protein or glycoprotein having a marker whose amino acid sequence has more than 70% 15 homology, preferably more than 85% homology, more preferably is identical with SEQ ID No. 1.

The present invention is furthermore also related to an inhibitor with an SDS-page molecular weight of typically 30 kDa being a protein or glycoprotein having 20 a marker whose amino acid sequence has more than 70% homology, preferably more than 85% homology, more preferably is identical with SEQ ID No. 1.

The present invention is furthermore also related to an inhibitor with an SDS-page molecular weight 25 of typically 10 kDa being a protein or glycoprotein having a marker whose amino acid sequence has more than 70% homology, preferably more than 85% homology, more preferably is identical with SEQ ID No. 2.

Advantageously, said markers are the end-30 terminal amino acid sequences of the protein or glycoprotein.

According to the invention, a marker of a

protein or glycoprotein means a specific amino acid sequence (or its corresponding nucleotide acid sequence) that is able to distinguish one protein family from another protein family.

5 The inhibitory effect towards xylan and/or arabinoxylan hydrolysing enzymes can be e.g. demonstrated by the endoxylanase method with AZCL arabinoxylan (cfr. infra). Likewise, the inhibitory effect towards  $\beta$ -glucan hydrolysing enzymes can be e.g. demonstrated by the  $\beta$ -  
10 glucanase method with AZCL  $\beta$ -glucan (cfr. infra).

The invention also relates to a method for obtaining said inhibitor from a micro-organism, such as a genetically modified micro-organism which expresses said inhibitors, from a plant, or from a plant material such as  
15 cereals, cereal grains, cereal flours or fractions 'thereof), by subjecting said plant, said plant material and/or said micro-organism to one or more extraction and/or fractionation steps.

Another aspect of the present invention is  
20 related to a method for genetically transforming a micro-organism, a plant or a plant material in order to obtain the expression of the inhibitor according to the invention wherein the micro-organism, the plant or plant material is genetically modified by the introduction of a genetic  
25 material encoding said inhibitor into the micro-organism, the plant or plant material and obtain its translation and expression by genetic engineering methods well known by the man skilled in the art.

The invention furthermore relates to  
30 processes aiming at changing, preferably reducing or increasing level of said inhibitor in a micro-organism, a plant or a plant material, by reducing or increasing the

expression of said inhibitor, by the methods well known by the man skilled in the art and/or by using molecules which are able to block the inhibitor activity or activate said inhibitor.

5 The invention furthermore relates to the obtained inhibitor, micro-organism, plant, plant material, and/or fractions thereof and to their use in different areas of food, feed and/or beverage technologies, such as improving malting and brewing, improving animal feedstuffs  
10 efficiency, baked and/or extruded cereal products (such as straight dough, sponge and dough and Chorleywood breads, breakfast cereals, different types of biscuits, pasta and noodles), improving the production of starch derived syrups, sorbitol, xylose and/or xylitol, improving wheat  
15 gluten-starch separation and production, maize processing, improving plant disease resistance, improving nutraceutical or pharmaceutical applications (such as maintaining the structure of dietary fiber material), and improving paper and pulp technologies.

20 The present invention will be described in details in the following description of a preferred embodiment without limiting the scope of the present invention.

25 Detailed description of the invention

During the course of their work dealing with the structure of arabinoxylans in Belgian white beers and in intermediates in the production process, the inventors unexpectedly found indications for inhibition of the  
30 xylanolytic barley malt system by wheat water extractables. This has not been reported before, although it has clearly been established that endogenous and exogenous  $\alpha$ -amylase

(Deponte, R., Parlamenti, T., Petrucci, V., Silano, V., & Tomasi, M., *Cereal Chemistry*, 1976, 53, 805; Buonocore, V., Petrucci, T., & Silano, V., *Phytochemistry*, 1977, 16, 811; Mundy, J., Hejgaard, J., & Svendsen, I., *Federation of Societies*, 1984, 167, 210; Silano, V.  $\alpha$ -Amylase inhibitors. In: *Enzymes and their Role in Cereal Technology*, J.E. Kruger, D. Lineback and C.E. Stauffer, (Eds). Am. Assoc. Cereal Chem., St. Paul (MN), 1987, 141) and protease (Birk, Y., *Methods Enzymology*, 1976, 45, 723; Lawszkowski, M., & Kato, I., *Annual Review of Biochemistry*, 1980, 49, 593) inhibitors are present in cereal grains.

Indeed, when one measured the solubilization of arabinoxylans during brewing with barley malt and unmalted wheat, with the objectives (1) to relate enzymic activities of the starting materials with the arabinoxylan contents of corresponding worts, and (2) to investigate in which way wheat interferes with the solubilization of arabinoxylans during brewing, there was evidence for the presence of xylanase inhibitors in wheat. This was indeed observed when one compared the solubilization of arabinoxylans and the release of free xylose (Xyl) in wort prepared with 60% malt and 40% wheat with that in a 100% malt wort.

Under certain experimental conditions, the addition to the wort of a xylanase of microbial origin clearly improved arabinoxylan solubilization during wort preparation.

#### Examples

#### 30 Materials

$\beta$ -D-Allose,  $\beta$ -mercaptoethanol, p-nitro-phenyl- $\beta$ -D-xylopyranoside and Trizma base (reagent grade,

tris[hydroxymethyl]amino-methane) were obtained from Sigma, St-Louis, MO, USA. Azurine-crosslinked (AZCL) wheat arabinoxylan (Xylazyme arabinoxylan tablets), AZCL and Xylanase M4 from Aspergillus niger was from Megazyme, Bray, 5 Ireland. Microbial xylanases from the micro-organisms *Bacillus substillis*, *Trichoderma viride* and *Aspergillus niger* were obtained from NV Puratos, Groot-Bijgaarden, Belgium. Buffer A was: 0.025 M sodium acetate, pH 4.7; Buffer B was: 0.025 M sodium maleate, pH 6.0; Buffer C was: 10 0.025 M sodium phosphate pH 6.0; Buffer D was: 0.250 mM sodium acetate pH 5.0; Buffer E was: 0.025 M sodium acetate pH 5.0.

Barley malt samples were supplied by Cargill Malt Division, Herent (Belgium). The inventors used a two- 15 rowed winter barley variety (Clarine) with a low endoxylanase activity and low water-extractable Xyl content, and two malts from a six-rowed winter barley variety Plaisant, with a high water-extractable Xyl content. Plaisant malt samples 1 and 2 had high and low 20 endoxylanase activities respectively. Wheat samples were from Amylum, Aalst (Belgium) and SAPSA SES SA, Jodoigne (Belgium). The inventors used Skirlou and Soissons with high and low water-extractable Xyl contents, respectively. Wheat germs were supplied by Ceres, Vilvoorde (Belgium). 25 Rye flour was from a mixture of Dutch rye varieties supplied by Meneba, Rotterdam (The Netherlands). Barley from the variety Clarine was supplied by Cargill, Malt Division, Herent (Belgium). Clarine barley, Plaisant 1 and Plaisant 2 barley malts, Skirlou and Soissons wheat 30 wholemeals were prepared either with the Tecator sample mill (Höganäs, Sweden) or for the brewing experiments with an EBC-approved laboratory mill (Analytica-EBC, Fourth

edition, Brauerei- und Getränke- Rundschau, Zurich, 1987). Soissons wheat flour was produced with a Bühler MLU-202 laboratory mill (Bühler, Uzwil, Switzerland, extraction yield 70%).

5

### Extracts

#### *BMWM1 WWM, and WG*

Samples (3.00 g) of ground barley malt and wheat or wheat germ (1.00 g) were suspended in buffer A (10.0 mL). After 15 min of vigorous shaking at room temperature, the suspensions were centrifuged (3,000 g, 15 min, 20°C). The resulting extracts are referred to as BMWM1 (barley malt wholemeal extract 1), WWM (wheat wholemeal extract), and WG (wheat germ extract).

15 *WF, RF, and BWM*

Samples of the appropriate flour or wholemeal (2.50 g) were suspended in buffer B (10.0 mL). After 15 min of vigorous shaking at room temperature, the suspensions were centrifuged (10,000 g, 15 min, 20°C) and the supernatants were filtered (0.45 µ). The resulting wholemeal extracts are referred to as WF (wheat flour extract), RF (rye flour extract), and BWM (barley wholemeal extract).

#### *BMWM2*

25 Samples (5.00 g) of ground barley malt were suspended in buffer B (10.0 mL). After 15 min of vigorous shaking at room temperature, the suspensions were centrifuged (10,000 g, 15 min, 20°C) and the supernatants were filtered (0.45 µ). The resulting wholemeal extracts 30 are referred to as BMWM2 (barley malt wholemeal extract 2).

Methods*Determination of Xyl content*

Extraction and hydrolysis procedures were as described by Cleemput et al (Cleemput, G., Roels, S.P., van Oort, M., Grobet P.J. & Delcour, J.A., *Cereal Chemistry*, 1993, 70, 324), with heating (130°C) of samples of whole meal (wheat and barley malt) for 5 hours to eliminate enzyme activity prior to extraction. Worts were analysed in the same way as the water-extracts of the whole meal flours. Free Xyl was determined by omitting the hydrolysis step prior to alditol acetate preparation. Alditol acetate samples (1 $\mu$ l) (Englyst, H.N. & Cummings J.H., *Analyst*, 1984, 109, 937) were separated at 225°C on a Supelco SP-2380 column (30 m, 0.32 mm ID, 0.2  $\mu$ m film thickness) and detected with a flame ionisation detector in a Chrompack 9011 Chromatograph (Middelburg, The Netherlands). Injection and detection temperatures were 275°C.  $\beta$ -D-Allose was used as internal standard. The arabinose (Ara) measured originated from both arabinoxylan and arabinogalactan making it impossible to calculate arabinoxylan levels as  $0.88 \times (\text{Ara} + \text{Xyl})$  (Cleemput, G., van Oort, M., Hessing, M., Bergmans, M.E.F., Gruppen, H., Grobet, P.J., Delcour, J.A., *Journal of Cereal Science*, 1995, 22, 73-84). Moreover, as in water-extracts of wheat wholemeal a substantial part of the galactose (Gal) does not stem from arabinogalactan, correction of Ara figures for arabinogalactan by assuming that the Gal/Ara ratio in arabinogalactan is 1.5 as known for wheat flours (Izydorczyk, M., Biliaderis, C.G. & Bushuk, W., *Cereal Chemistry*, 1991, 68, 139-144) was equally impossible. In what follows, therefore, the Xyl figures are used as a relative measure for arabinoxylans levels. In a similar

way, the increase in Xyl levels during brewing is indicative of arabinoxylan solubilization during brewing.

Measurement of endoxylanase (EC 3.2.1.8) activity and  
5 inhibition thereof

Extracts (1.0 mL) BMWM1 and WWM (cfr. supra) were incubated for 5 min at 50°C, before adding an AZCL-xylan tablet (Megazyme). The incubation was then continued for 60 min at 50°C. The reaction was terminated by adding  
10 1% (w/v) Trizma base (10.0 mL) and vigorous vortex stirring. After 5 min at room temperature, the tubes were shaken vigorously and the contents filtered through a Whatman N°1 filter. The absorbance was measured at 590 nm against a control, which was prepared by incubating the  
15 extract without the substrate tablet for 60 min at 50°C. The substrate tablet was added after adding 1% (w/v) Trizma base to the extract. The activity was expressed as the difference in the absorbance at 590 nm between the sample and control and expressed per gram dry malt ( $\Delta A_{590}/g$ ).

20 The endoxylanase activity of 0.6 mL BMWM1 (cfr. supra) and 0.4 mL buffer A was compared with the activity of 0.6 mL BMWM1 to which 0.4 mL WWM was added. In some cases, the WWM was boiled for 30 min and centrifuged (3,000 g, 15 min, 20°C) prior to addition.

25 In the evaluation of the inhibition of microbial enzymes by extracts from different cereals the following procedure was used. Extracts (WF, WG, RF, and BWM) or boiled (30 min, 100°C) and centrifuged (10,000 g, 15 min, 20°C) extracts (250  $\mu$ L) were preincubated for 30  
30 min at room temperature with 250  $\mu$ L of appropriately diluted microbial xylanase solution, the xylazyme tablet was added and the mixture was incubated for 60 min at 50°C.

The remainder of the procedure was as described above with addition of 5.0 mL 2% (w/v) Trizma base instead of 10.0 mL 1% (w/v) to terminate the reaction.

5 Measurement of  $\beta$ -glucanase (EC: 3.2.3.73) activity and inhibition thereof

Extracts (WF, RF, and BWM) or boiled (30 min, 100°C) and centrifuged (10,000 g, 15 min, 20°C) extracts (450  $\mu$ L) were preincubated for 30 min at room temperature

10 with 50  $\mu$ L of BMWM2, the  $\beta$ -glucazyme tablet was added and the mixture was incubated for 60 min at 40°C. The remainder of the procedure was as described above with addition of 5.0 mL 2% (w/v) Trizma base instead of 10.0 mL 1% (w/v) to terminate the reaction.

15

*Brewing*

Worts were prepared in duplicate according to the EBC method (Analytica-EBC, Fourth edition, Brauerei- und Getränke- Rundschau, Zurich, 1987). For the 100% barley 20 malt worts, 50.0 g barley malt was used and for the worts with 40% wheat, 30.0 g barley malt and 20.0 g wheat. Worts were centrifuged for 15 min at 2,000  $\times$  g (room temperature). The spent grains were washed (150 mL) and the washings were added to the worts.

25 The *Bacillus subtilis* endoxylanase was added to the water (46°C) before mixing with 60% Clarine malt and 40% Soissons or Skirlou wheat. The level of endoxylanase added to worts ( $0.867 \Delta A_{590}/g$ ) was equal to that needed to increase the endoxylanase activity of Clarine malt (0.750 30  $\Delta A_{590}/g$ ) to the level in Plaisant 1 malt ( $1.617 \Delta A_{590}/g$ ).

All analyses described above were carried out

at least in duplicate and the mean values are presented. The experimental error (E.E.) was calculated from the difference (in %) between the individual and the mean values.

5

#### SDS-Polyacrylamide gel electrophoresis and iso-electrofocusing

The molecular weight of the purified inhibitor was determined by SDS-polyacrylamide gel electrophoresis (SDS-page) on 20% polyacrylamide gels under reducing ( $\beta$ -mercaptoethanol, 1%) or non-reducing conditions with the PhastSystem unit (Pharmacia, Uppsala, Sweden), according to the method of Laemmli, U.K. (Nature, 1970, 227, 680-685). The gels were silver stained according to the instructions of the manufacturer (Pharmacia, Development Technique file N° 210). Low molecular weight markers were  $\alpha$ -lactalbumin (14,400 Da); trypsin inhibitor (20,100 Da); carbonic anhydrase (30,000 Da); ovalbumin (43,000 Da); albumin (67,000 Da); phosphorylase b (94,000 Da). The iso-electric point of the inhibitor was determined with the PhastSystem unit using polyacrylamide gels containing ampholytes (pH 3-9) and with appropriate standards (Pharmacia calibration kits, pI 3.5 - 9.3). The proteins were silver stained (cfr. supra).

25

#### N-terminal amino acid sequencing of proteins.

The sequences of the N-terminal amino acids were determined with an Applied Biosystems model 477 A gas-phase sequencer, connected on line with an 120 A PTH analyser (Perkin Elmer, Belgium).

Evidence for the presence of endoxylanase inhibitors in wheat

Barley malt and wheat Xyl levels and arabinoxylan hydrolysing activities are listed in Table I.

5 Xyl levels in the 100% malt worts (Table II) varied from 0.41 to 0.62% (all analytical data expressed as percentage of dry matter). The Xyl levels in the worts with 40% wheat varied from 0.35 to 0.61% (Table III).

In the worts with 40% wheat, the inventors 10 used 60% barley malt. Comparison of the increase in Xyl during brewing using 60% malt with 60% of the Xyl increase using 100% barley malt showed a reduction of 12 to 58% (Tables II and III). This suggested that the endoxylanases from barley malt were inhibited in the presence of wheat or 15 that the wheat arabinoxylans are a less suitable substrate for malt endoxylanases. Malting breaks down barley cell walls and renders them more accessible for enzymes (Selvig, A., & Aarnes, H., *Journal of the Institute of Brewing*, 1986, 92, 185).

20

Free Xyl levels in wort

The levels of free Xyl in 100% malt worts varied from 0.046 to 0.076% and in the worts with 40% wheat from 0.025 to 0.040% (Table IV). The difference between the 25 levels of the released free Xyl was 0.032 to 0.044% for the 100% malt worts and 0.015 to 0.020% for the worts with 40% wheat. The reduction in free Xyl release compared with 60% of the free Xyl release with the 100% barley malt wort varied from 1 to 32% (Tables II and IV). The use of the 30 *Bacillus subtilis* endoxylanase did not result in an increase of free Xyl. The free Ara levels did also not increase. The endoxylanase, therefore, had no side  $\beta$ -D-

xylosidase and  $\alpha$ -L-arabinofuranosidase activities.

The reduction of the endoxylanase induced increase in Xyl or arabinoxylan solubilization as a result of the use of wheat in conjunction with barley malt was 5 more obvious than the reduction of the release of free Xyl. For this reason one focused on the inhibition of the barley malt endoxylanases by a wheat component.

*Malt xylanolytic system inhibition by wheat extracts*

10 In Figure 1 the reduction of the endoxylanase activity of BMWM1, when WWM instead of buffer A was added, is given. The reduction of endoxylanase activity varied from 26 to 58 %. The reduction of endoxylanase activity of barley malt wholemeal extracts (BMWM1) was obtained by 15 addition of wheat wholemeal extracts (WWM) instead of buffer. The figure 1 represents the results obtained with unboiled (□) and boiled extracts (■). (a) Clarine malt + Soissons wheat, (b) Clarine malt + Skirlou wheat, (c) Plaisant 1 malt + Soissons wheat, (d) Plaisant 1 malt + 20 Skirlou wheat, (e) Plaisant 2 malt + Soissons wheat, (f) Plaisant 2 malt + Skirlou wheat.

A higher reduction was observed in case of cv. Skirlou than with cv. Soissons. This was in line with the higher reduction of Xyl increase during brewing with 25 cv. Skirlou than with cv. Soissons (see Table III). The higher water extractable Xyl content of cv. Skirlou than for cv. Soissons implied that the lower susceptibility of the wheat substrate during brewing may cause the reduced solubilization. When boiling WWM, almost all of the 30 inhibition disappeared. The inhibitor seemed to be thermolabile and the inventors concluded that it therefore may be of proteic nature. However, it was considered

unlikely that one dealt with a protease as the protease activity from the malt is many times higher than the protease activity of wheat. The major part of the reduced activity was apparently not caused by the wheat 5 arabinoxylans because the thermal treatment did not change the arabinoxylan concentration of the wheat extract. Whether the wheat inhibitor was active against the endogenous barley malt endoxylanases or exogenous endoxylanases was unclear.

10

Brewing with *Bacillus subtilis* xylanase: a solution for poor arabinoxylan solubilisation in barley malt - wheat wholemeal brewing

The *Bacillus subtilis* endoxylanase, one of 15 the microbial enzymes that was relatively little inhibited under the experimental conditions of Figure 2, increased the Xyl levels present in wort. In comparison with the same worts made without endoxylanase addition, 94% more Xyl solubilization using cv. Soissons as wheat adjunct and 179% 20 more Xyl solubilization using cv. Skirlou as wheat adjunct was obtained. The *Bacillus subtilis* endoxylanase apparently solubilizes more arabinoxylan from the wheat variety Skirlou than from the wheat variety Soissons (Table IV).

25 Purification of xylanase inhibitor from wheat flour

Soissons wheat flour (2.0 kg) was suspended in 10.0 L 0.1% (w/v) ascorbic acid. The suspension was mixed overnight at 7°C and centrifuged (7°C, 10,000 g, 30 min). To the supernatant 2.0 g/l  $\text{CaCl}_2$  was added and the pH 30 was raised to 9.0 by addition of 2.0 M NaOH. The extract was left overnight at 7°C and centrifuged (7°C, 10,000 g, 30 min). The pH of the extract was adjusted to 5.0 with 2.0

M HCl and the extract was pumped over a cation exchanger (SP Sepharose Fast Flow, 50 × 50 mm, Pharmacia). The column was equilibrated with buffer C (200 mL) and a protein fraction was eluted with 200 mL 0.5 M NaCl. This eluate was 5 diluted 5 times, the pH adjusted to 5.0 as above and cations were exchanged (SP Sepharose Fast Flow, 26 × 100 mm, Pharmacia). The column was equilibrated with buffer C (200 mL) and after a linear salt gradient from 0 to 0.5 M NaCl (800 mL), fractions of 10 mL were, after desalting (PD 10 column, Pharmacia), assayed for endoxylanase inhibition 10 using the cited xylazyme method with eluate instead of cereal extract and appropriately diluted Xylanase M4 from *Aspergillus niger*. The fractions with inhibition activity were collected, dialyzed against dionised water (7°C, 15 overnight) and lyophilised. The lyophilised material was dissolved in buffer D (6.0 mL) and separated on a Sephadryl S100 column (26 × 670 mm, Pharmacia) eluted with the same buffer. Fractions of 2.5 mL were collected and assayed for inhibitor activity. The active fractions were collected, 20 dialyzed as above and lyophilised. The lyophilised material was dissolved in buffer E (6 mL) and cation exchanged (Mono S HR 5/5, Pharmacia) with the same buffer. Fractions eluted in a salt gradient (0 to 0.5 M NaCl) were collected and assayed for xylanase inhibition as above. In this way, 25 we obtained a fraction (1 mL) of the inhibitor which migrated as a single protein band on SDS-PAGE. It had an apparent molecular weight of ca. 40 - 43 kDa. Following reduction with  $\beta$ -mercaptoethanol, two additional SDS-PAGE bands of molecular weights of typically 30 and 10 kDa are 30 detected.

*N-terminal amino acid sequencing of endoxylanase inhibitor*

The  $\beta$ -mercaptoethanol reduced inhibitor protein and/or glycoprotein fractions were subjected to SDS-page, blotted and N-terminal amino acid sequenced.

5 The N-terminal amino acid sequence of the ca. 40 - 43 kDa band (SEQ ID No.01) was: Lys-Gly-Leu-Pro-Val-Leu-Ala-Pro-Val-Thr-Lys-Xaa-Thr-Ala wherein Xaa being preferably Asp. This sequence has not been reported before.

10 The above cited ca. 30 kDa band also has the above described typical N-terminal amino acid SEQ ID No.1, while the N-terminal amino acid sequence of the ca. 10 kDa band is typically as follows: SEQ ID No. 2: Xaa-Ala-Pro-Val-Ala-Lys-Met-Val-Leu-Pro-Val-Ala-Met-Lys-Glu-Xaa-Val, wherein the first Xaa being preferably Ser, Phe, or Gly, and 15 wherein the second Xaa is unidentified. This sequence has not been described before.

Inhibition of different microbial endoxylanases by endoxylanase inhibitors from wheat and other cereals

20 In Figure 2 the inhibition of different microbial xylanases in the presence of WF, RF, and BWM is shown. The reduction of the xylanase activity (%) when a cereal extract was added instead of the same extract boiled for 30 min is given. Under the experimental conditions, the 25 highest reduction was found for the mixture of three xylanases from *Trichoderma reesei* (82 to 94%) the lowest for the xylanases from *Bacillus subtilis* (24 to 39%).

The reduction of microbial endoxylanase activity was obtained by addition of cereal extracts (WF, 30 RF, and BWM) instead of boiled cereal extracts. The figure 2 represents the results obtained with wheat flour (■),

rye flour (□) and barley whole meal (■). Microbial xylanases: (a) Mixture of three xylanases from *Trichoderma reesei*, (b) Xylanase M4 from *Aspergillus niger*, (c) Xylanase from *Bacillus subtilis*, (d) Mixture of three xylanases from *Bacillus subtilis*, (e) Xylanase from *Aspergillus niger*, (f) Mixture of five xylanases from *Aspergillus niger*, (g) Mixture of five xylanases from *Aspergillus niger*. Under the experimental conditions, WG reduced the activity of xylanase M4 from *Aspergillus niger* with ca. 80 %.

Inhibition of barley malt  $\beta$ -glucanase by inhibitors from wheat and other cereals

In Figure 3 the inhibition of malt  $\beta$ -glucanase in the presence of WF, RF, and BWM is shown. The reduction of the  $\beta$ -glucanase activity (%) when a cereal extract was added instead of the same extract boiled for 30 min is given. The reduction varied from 7 to 12%.

The reduction of  $\beta$ -glucanase activity of barley malt extracts (BMWM2) was obtained by addition of cereal extracts (WF, RF, BWM) instead of boiled cereal extracts. The figure 3 represents the results obtained with wheat flour (a), rye flour (b) and barley whole meal (c).

TABLE I. Water-extractible and Free Xylose Contents (% of Dry Matter) and Arabinoxylan Degrading Enzyme Activities of 3 Barley Malts and 2 Wheats\*.

	Water-extractable Xyl	Free Xyl	Endoxylanase ( $\Delta A_{590}/g$ )	$\beta$ -D-Xylosidase (U/g)
Barley Malt				
Clarine	0.29	0.014	0.750	0.286
Plaisant 1	0.41	0.031	1.617	0.331
Plaisant 2	0.40	0.013	0.607	0.299
Wheat				
Soissons	0.27	0.005	0.115	0.054
Skirlou	0.52	0.003	0.205	0.053
E.E. < 7%	E.E. < 4%	E.E. < 9%	E.E. < 6%	

\*Xyl = xylose; Water-extractable Xyl = total - free xylose water-extract; E.E. = experimental error.

TABLE II Xylose and Free Xylose Levels (% of Dry Matter) in Wort and Levels of Increase in Xylose and Free Xylose (% of Dry Matter) during Brewing with 100% Barley Malt\*.

Barley Malt	Xyl Wort	Xyl Increase	Free Xyl Wort	Free Xyl Increase
Clarine	0.41	0.15	0.046	0.032
Plaisant 1	0.62	0.26	0.075	0.044
Plaisant 2	0.51	0.09	0.049	0.035

E.E. < 6%

E.E. < 7%

\*Xyl = xylose; Xyl Wort = total - free xylose wort; Xyl Increase = total xylose wort - total xylose water-extract barley malt; Ara = arabinose; E.E. = experimental error.

TABLE III

Xylose Levels (% of Dry Matter) in Grains and Corresponding Worts Prepared with 60% Barley Malt and 40% Wheat. Increase in Xylose Levels (% of Dry Matter) during Brewing and Effect of Addition of *Bacillus subtilis* Endoxylanase. Difference (%) with 60% of the Increase of Xylose Levels in Case of a 100% Malt Wort\*.

Barley Malt (60%) + Wheat (40%)	Xyl Grains	Xyl Wort	Xyl Increase	Difference 100%	Malt Wort
Clarine + Soissons	0.29	0.35	0.08	-12	
Clarine + Skirlou	0.41	0.46	0.07	-27	
Plaisant 1 + Soissons	0.35	0.44	0.10	-36	
Plaisant 1 + Skirlou	0.48	0.53	0.06	-58	
Plaisant 2 + Soissons	0.35	0.39	0.06	-28	
Plaisant 2 + Skirlou	0.48	0.51	0.05	-40	
Clarine + Soissons + BSX	0.31	0.45	0.16	93	
Clarine + Skirlou + BSX	0.44	0.61	0.19	130	

E. E. &lt; 8%

E. E. &lt; 7%

\*Xyl = xylose; Xyl Grains =  $0.6 \times (\text{total} - \text{free xylose water-extract barley malt}) + 0.4 \times (\text{total} - \text{free xylose water-extract wheat})$ ; Xyl wort = total - free xylose wort; Xyl Increase = total xylose wort -  $[0.6 \times (\text{total xylose water-extract barley malt}) + 0.4 \times (\text{total xylose water-extract wheat})]$ ; Difference 100% Malt Wort =  $[100 \times (\text{increase xylose wort from 60% malt and 40% wheat}) / (\text{xylose increase wort from 100% malt})] - 100$ ; BSX = *Bacillus subtilis endoxylanase*; E.E. = experimental error.

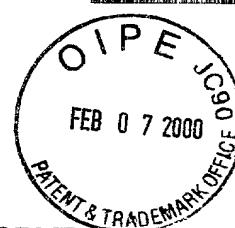
TABLE IV

Release of Free Xylose during Brewing with 60% Barley Malt and 40% Wheat. Effect of Addition of *Bacillus subtilis* Endoxylanase. Difference (%) with 60% of the Release of Xylose in Case of a 100% Malt Wort\*.

Wheat (40%)	Free Xyl			Malt Wort
	Grains	Wort	Release	
Barley Malt (60%) +				
Clarine + Soissons	0.010	0.025	0.015	-20
Clarine + Skirlou	0.010	0.029	0.019	-1
Plaisant 1 + Soissons	0.021	0.039	0.018	-32
Plaisant 1 + Skirlou	0.020	0.040	0.020	-24
Plaisant 2 + Soissons	0.010	0.029	0.019	-9
Plaisant 2 + Skirlou	0.009	0.029	0.020	-5
Clarine + Soissons + BSX	0.010	0.026	0.015	-19
Clarine + Skirlou + BSX	0.010	0.029	0.019	-2
			E.E. < 8%	E.E. < 7%

\*Xyl = xylose; Difference 100% Malt Wort =  $[100 \times (\text{xylose release wort from 60% malt and 40% wheat}) / (\text{xylose release wort from 100% malt})] - 100$ ; BSX = *Bacillus subtilis* endoxylanase;

E.E. = experimental error.



P. KUL. 02/03 (20)

DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATIONAttorney Docket No: VANM131.001APCFirst Named Inventor: winok DebyslerComplete if known: Serial No: 09/403,625 Filing Date: \_\_\_\_\_Group Art Unit: Unknown Examiner: Unknown

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled INHIBITORS OF CELLULLOLYTIC, XYLANOLYTIC AND B-GLUCANOLYTIC ENZYMES,

specification of which:  is attached hereto or  was filed on May 4, 1998 (Int'l) assigned application Serial No. 09/403,625, and was amended on (if applicable) filed as International Application No. PCT/EP98/02590.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, S. 1.56(a).

I hereby claim foreign priority benefits under 35 U.S.C. 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below any foreign application for patent or inventor's certificate or of any PCT international application having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s):

<u>97870060.7</u> (Number)	<u>Europe</u> (Country)	<u>April 30, 1997</u> (Month/Day/Year Filed)	<u>Priority Claimed</u>	<u>Certified Copy</u>		
			<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No	<u>Attached</u>	<input type="checkbox"/> Yes
<u>(Number)</u>	<u>(Country)</u>	<u>(Month/Day/Year Filed)</u>	<input type="checkbox"/> Yes	<input type="checkbox"/> No	<input type="checkbox"/> Yes	<input type="checkbox"/> No

I hereby claim the benefit under 35 U.S.C. 119(e) of any United States provisional application(s) listed below:

Application No: \_\_\_\_\_ Filing Date: \_\_\_\_\_

I hereby claim the benefit under 35 U.S.C. 120 of any United States application(s), or 365(c) of any PCT international application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C 112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

US Parent Application or PCT  
Parent Number

Parent Filing Date

Parent Patent Number  
(if applicable)

And I hereby appoint

KNOBBE, MARTENS, OLSON & BEAR, LLP  
620 Newport Center Drive, 16th Floor  
Newport Beach, CA 92660

attorneys with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent Office connected therewith.

Please direct all future correspondence in connection with this application to the attention of

DANIEL E. ALTMAN

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor: WINOK DEBYSER WINOK DEBYSER

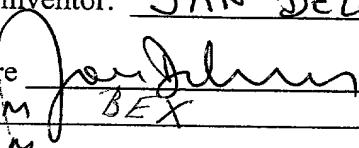
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